

Slovenská poľnohospodárska univerzita v Nitre
Fakulta biotechnológie a potravinárstva
Fakulta agrobiológie a potravinových zdrojov

V. Vedecká konferencia doktorandov

s medzinárodnou účasťou

pri príležitosti Európskeho týždňa vedy



**FAKULTA
BIOTECHNOLÓGIE
A POTRAVINÁRSTVA**



**FAKULTA
AGROBIOLOGIE
A POTRAVINOVÝCH
ZDROJOV**

Nitra, 26. november 2010

V. Vedecká konferencia doktorandov

s medzinárodnou účasťou pri príležitosti Európskeho týždňa vedy

Vedeckí garanti:

prof. Ing. Ján Jančovič, PhD.
doc. RNDr. Dana Urminská, CSc.

Organizační garanti:

doc. Ing. Janette Musilová, PhD.
Ing. Ľuboš Vozár, PhD.
Ing. Henrieta Arpášová, PhD.
doc. Ing. Kamil Hudec, PhD.
Ing. Silvia Melicháčová, PhD.
Ing. Mária Timoracká, PhD.
doc. Ing. Janette Musilová, PhD.
Ing. Ľuboš Vozár, PhD.

Zostavili:

Lektorovali:

Ing. Henrieta Arpášová, PhD.
RNDr. Daniel Bajčan, PhD.
Ing. Marek Bobko, PhD.
doc. Ing. Marcela Capcarová, PhD.
doc. Ing. Margita Čanigová, CSc.
prof. Ing. Juraj Čuboň, CSc.
Ing. Viera Ducková, PhD.
Ing. Katarína Fatrnecová Šramková, PhD.
prof. RNDr. Zdenka Gálová, CSc.
Ing. Miroslav Habán, PhD.
Ing. Marta Habánová, PhD.
doc. Ing. Peter Haščík, PhD.
doc. Ing. Kamil Hudec, PhD.
Ing. Milan Chňapek, PhD.
doc. Ing. Peter Chrenek, DrSc.
prof. Ing. Ján Jančovič, PhD.
doc. RNDr. Klaudia Jomová, PhD.
doc. Ing. Miroslava Kačániová, PhD.
Ing. Miriam Kádasi Horáková, PhD.
doc. Ing. Adriana Kolesárová, PhD.
Ing. Miroslav Kročko, PhD.
doc. Ing. Ladislav Lagin, CSc.
Ing. Ladislav Lahučký, PhD.

Doc. Mgr. Ing. Peter Lazor, PhD.
MVDr. Ľubomír Lopašovský, PhD.
doc. Ing. Norbert Lukáč, PhD.
Ing. Ján Mareček, PhD.
doc. MVDr. Peter Massanyi, PhD.doc.
Ing. Silvia Melicháčová, PhD.
Ing. Andrea Mendelová, PhD.
RNDr. Juraj Mišík
doc. Ing. Janette Musilová, PhD.
Mgr. Ing. Adriana Pavelková, PhD.
Ing. Anežka Poláková, PhD.
Ing. Radovan Stanovič, PhD.
doc. Ing. Peter Strapák, PhD.
Ing. Eva Szabová, PhD.
doc. Ing. Dana Tančinová, PhD.
Ing. Mária Timoracká, PhD.
doc. Ing. RNDr. Tomáš Tóth, PhD.
Ing. Pavol Trebichalský, PhD.
Ing. Ladislav Varga, PhD.
Ing. Vladimír Victoris, PhD.
Ing. Martin Vivodík, PhD.;
Ing. Ľuboš Vozár, PhD.
Ing. Lucia Zeleňáková, PhD.

© Slovenská poľnohospodárska univerzita v Nitre

Zborník neprešiel jazykovou úpravou, za obsah sú zodpovední autori.

Schválil rektor Slovenskej poľnohospodárskej univerzity v Nitre dňa 25. 10. 2010 ako zborník z vedeckej konferencie

ISBN 978-80-552-0471-0

SLEDOVANIE ANTIBIOTICKEJ REZISTENCIE BAKTÉRIÍ Z ČELADE ENTEROBACTERIACEAE IZOLOVANÝCH Z REKTÁLNYCH VÝTEROV OVIEC, CECKOV A Mlieka

MONITORING OF ANTIBIOTIC RESISTANCE OF BACTERIA OF *ENTEROBACTERIACEAE*
GENERA ISOLATED FROM SHEEP RECTAL SWABS, TEAT AND MILK

Lukáš HLEBA, Miroslava KAČÁNIOVÁ, Jaroslav POCHOP

Abstrakt: Cieľom práce bolo zistiť antibiotickú rezistenciu čelade *Enterobacteriaceae* izolovanej z rektálnych výterov oviec, ceckov a mlieka z konvenčného chovu zo Slovenska. Kultivácia prebiehala na selektívnom agare MacConkey určený pre čelad' *Enterobacteriaceae*. Test antibiotickej citlivosti prebiehal na agare ISO sensitest agar s antibiotikami: Enrofloxacin (ENR 5), Tetracyklín (TE 30), Streptomycín (S 10) a Chloramphenicol (C 30). Pre identifikáciu druhov čelade *Enterobacteriaceae* sme použili identifikačné agary (Chromogénny agar pre koliformné baktérie, Hajnov agar, XLD agar) a pre biochemickú identifikáciu ENTERO test 24. V našej štúdií sme zistili, že najvyššia rezistencia druhov čelade *Enterobacteriaceae* bola (11,11 %) na Streptomycín a Chloramphenicol pri izolátoch z ceckov. Najvyššia rezistencia z izolátov mlieka bola v prípade Chloramphenicolu (5,88 %). V izolátoch z rektálnych výterov oviec bola najvyššia rezistencia čelade *Enterobacteriaceae* na Tetracyklín (3,70 %). Vo všetkých izolátoch však prevládala vysoká citlivosť ku všetkým použitým antibiotikám. Najvyššiu citlivosť mala čelad' *Enterobacteriaceae* na Enrofloxacin (100 %) zo všetkých izolátov. Pri Streptomycíne bola citlivosť 100 % v prípade izolátov z rektálnych výterov a mlieka. Pri Tetracyklíne bola rezistencia 100 % v prípade izolátov z ceckov a mlieka a pri Chloramphenicole bola citlivosť 100 % v prípade izolátov z rektálnych výterov. Z čelade *Enterobacteriaceae* sme pomocou identifikačných agarov a biochemických testov zistili tieto druhy: *Escherichia coli*, ktorá bola rezistentná k Tetracyklínu a Chloramphenicolu, *Serratia plymuthica*, ktorá bola rezistentná k Chloramphenicolu, *Citrobacter freundii*, ktorý bol rezistentný k Streptomycínu a Klebsiella pneumoniae, ktorá bola rezistentná k Chloramphenicolu. Z výsledkov vyplýva, že baktérie môžu prenášať svoje gény rezistencie nielen vertikálne ale aj horizontálne. Výsledky taktiež poukazujú na to, že gény pre antibiotickú rezistenciu neprechádzajú len tráviacim traktom, kde sa baktérie vyskytujú vo veľkej miere ale prechádzajú aj do finálnych produktov ako napr. mlieko. Na základe výsledkov môžeme konštatovať, že ľudský faktor zohráva významnú úlohu pri prenose génov rezistencie z človeka na zvieratá.

KLúčové slová: antibiotická rezistencia, *Enterobacteriaceae*, ovca, rektus, cecky, mlieko

Abstract: The aim of this study was to determine antibiotic resistance of *Enterobacteriaceae* genera isolated from sheep rectal, teat and milk from conventional livestock from Slovakia. The cultivation was done on a selective agar for *Enterobacteriaceae* (MacConkey). Antibiotic susceptibility testing was done on the ISO sensitest agar with these antibiotics: Enrofloxacin (ENR 5), Tetracycline (TE 30), Streptomycin (S 10) and Chloramphenicol (C 30). For identification of strains of *Enterobacteriaceae* genera, we used identification agar (Chromogenic coliform agar, Triple Sugar Iron agar, XLD agar) and for biochemical identification ENTERO test 24. In our study, we determined that the highest resistance of *Enterobacteriaceae* strains was to Streptomycin and Chloramphenicol (11.11 %) in isolates from sheep teat. The highest resistance was to Chloramphenicol (5.88 %) from isolates of milk. In the isolates from rectal swabs of sheep was the highest resistance to Tetracycline (3.70 %). In the all isolates, however, there was a high susceptibility to all antibiotics used. The highest susceptibility has *Enterobacteriaceae* genera to Enrofloxacin (100 %) from to all of isolates. Susceptibility to Streptomycin was 100 % from the isolates of rectal swab and milk. To Tetracycline 100 % in the isolates from teat and milk and to Chloramphenicol was susceptibility 100 % from isolates of rectal swabs of sheep. From these genera, we identified by identifying agars and biochemical tests: *Escherichia coli* resistant to Tetracycline and Chloramphenicol, *Serratia plymuthica* resistant to Chloramphenicol, *Citrobacter freundii* resistant to Streptomycin and *Klebsiella pneumoniae* resistant to Chloramphenicol. The results show that the bacteria can transfer resistance genes and their spread not only in vertical but also horizontal. Results also confirm that antibiotic resistance not only through the digestive tract of animals but also in their final product such as milk. It is also possible the results show that human factor plays important role in the transfer resistant genes from human to animal.

Key words: antibiotic resistance, *Enterobacteriaceae*, sheep, rectal, teat, milk

INTRODUCTION

Antibiotic resistance is significant health, social and economic problem at this time. Antibiotic resistance of bacteria is biological risk, which it is increase morbidity and mortality of animal and human (EFSA, 2008). The most technologies in the production and food processing reduced the incidence of pathogens including resistant bacteria to antibiotics. Experimental monitoring confirmed that the treatment of food technology based on damage to cell membranes and enzymes may help to generate and transfer of antibiotic resistance (Lado a Yousef, 2002).

Kharazmi et al., 2002; Mc Mahon et al., 2007). The health safety of foods, including milk, it is an integral part of consumers policy and health (Bíreš, 2004). Milk is a suitable substrate for the growth of many pathogenic and toxicogenic microorganisms which may be the cause of foodborne diseases that can endanger the health of the consumers (Bobková et al., 2008).

In our study, we followed antibiotic resistance of *Enterobacteriaceae* genera isolated from rectal of sheep, teat and milk from once conventional livestock from Slovakia.

MATERIAL AND METHODS

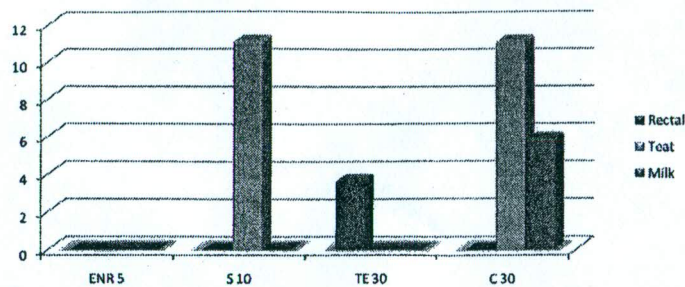
Antibiotics resistance study was done on *Enterobacteriaceae* genera isolated from rectal swabs of sheep, teat and milk. The bacterial strains were isolated from rectal swabs, teat, milk and collected with a kit containing the swab (Copan Inovation, Brescia) and the transport in medium to laboratory (Department of Microbiology, SUA in Nitra). For cultivation of *Enterobacteriaceae* MacConkey agar (Biomark, Pune) was used. Cultivation of these microorganisms was done during 24 hours for *Enterobacteriaceae* at 37 °C. The pure colonies were recultivation at the same conditions. The inoculum of *Enterobacteriaceae* was prepared by suspending of colonies from agar plates and the suspension was adjusted to equal a 0,5 McFarland standard. The sensitivity of all isolates was tested against: Streptomycin (S 10) 10 µg.disk⁻¹, Tetracycline (TE 30) 30 µg.disk⁻¹, Enrofloxacin (ENR 5) 5 µg.disk⁻¹ and Chloramphenicol (C 30) 30 µg.disk⁻¹. We used disk diffusion methods (according to EUCAST – European committee on antimicrobial susceptibility testing) (EUCAST). The incubation of strains with antibiotic disks was done on the ISO Sensitest agar (Biolife, Italiana) at the 37 °C. The interpretation of inhibition zones around the disk was according to BSAC version 8 January 2009 British Society for Antimicrobial Chemotherapy. The inhibition zones were controlled with reference strains *Escherichia coli* ATCC 25922. The basic differentiation of strains of *Enterobacteriaceae* genera was done on the identification agars like MacConkey agar (Biomark, Pune), Chromogenic coliform agar (Biolife, Italiana), Triple Sugar Iron agar (Biolife, Italiana) and XLD agar (Biolife, Italiana). Biochemical identification of strains of *Enterobacteriaceae* genera was done by ENTEROtest 24 (Pliva, Lachema). For the evaluation of biochemical test we used identification program TNW Lite 7.0 (Pliva, Lachema).

RESULTS AND DISCUSSION

We study antibiotic resistance in commensal *Enterobacteriaceae* genera, which are considered a potential reservoirs of resistant genes in environment. This reservoirs of resistant bacteria provide a potential source for resistant genes transfer between bacteria as well as an environment and food products. Therefore, identifying these reservoirs and mechanisms of persistence will be a key to reducing the load of resistant bacteria in everywhere.

In our study, we determined that the highest resistance of *Enterobacteriaceae* strains was to Streptomycin and Chloramphenicol (11.11%) from isolates of sheep teat. From the isolates of milk was the highest resistance to Chloramphenicol (5.88%). To the Tetracycline was (3.70%) the highest resistance of *Enterobacteriaceae* genera from isolates of rectal swabs of sheep. The susceptibility to Enrofloxacin was 100% to all of used antibiotics. To the Streptomycin was 100% susceptibility from isolates of rectal swabs and sheep milk. The susceptibility to Tetracycline was 100% from isolates of teat and milk and to Chloramphenicol was 100% only from rectal swabs of sheep. Results are shown in the fig. 1 and tab. 1.

Reported results Solomakos et al. (2009), which detected antibiotic resistant in the milk coming from the Greek. Their results show higher resistance of *E. coli* to Ampicillin and Chloramphenicol. Farzana et al. (2009) determined that in Indian milk resistance was 100% to Chloramphenicol. The similar results like Farzana et al. (2009) determined Dupont et al. (1978) in their work too. The number of researchers, such as Lira et al. (2004), Picozzi et al. (2005), Caro et al. (2007), Čížek et al. (2007), which examined the antibiotic resistance of *E. coli*, respectively, *Enterobacteriaceae* genera isolated from milk have argued that the results of antibiotic resistance vary from study to study like our results.



Legend: ENR5 – Enrofloxacin, S 10 – Streptomycin, TE 30 – Tetracycline, C 30 – Chloramphenicol

Figure 1 Antibiotic resistance profil isolates of *Enterobacteriaceae* from sheep rectal, teat and milk

Table 1 Antibiotic resistance profil isolates of *Enterobacteriaceae* from sheep rectal, teat and milk

Antibiotics/Isolates from	Sheep rectal	Sheep teat	Sheep milk
ENR 5	0,00	0,00	0,00
S 10	0,00	11,11	0,00
TE 30	3,70	0,00	0,00
C 30	0,00	11,11	5,88

Of these samples isolated from rectal swabs, teat and sheep milk, we identified following strains: *Escherichia coli*, which was resistant to Tetracycline. *Serratia plymuthica*, which was resistant to Chloramphenicol. *Citrobacter freundii*, which was resistant to Streptomycin and *Klebsiella pneumoniae*, which was resistant to Chloramphenicol. In the tab. 2 are shown results of identification strains of *Enterobacteriaceae* genera in percentage.

Table 2 Identified resistant strains and percentage of identification by ENTERO test 24

Strains	Identification (%)	Resistance to
<i>Escherichia coli</i>	100	Tetracycline, Chloramphenicol
<i>Serratia plymuthica</i>	100	Chloramphenicol
<i>Citrobacter freundii</i>	96,24	Streptomycin
<i>Klebsiella pneumoniae</i>	92,99	Chloramphenicol

CONCLUSION

Use of antibiotics in livestock farming causes that more obligatory and facultative pathogens are resistant to various antibiotics used commercially. These pathogens microorganisms can cause many different infections or diseases, which are then difficult to treat. Our experiment results show that antibiotic were used in livestock farming or rearing were introduced into the external environment. The results show that the bacteria can transfer resistance genes and their spread not only in vertical but also horizontal. Results also confirm that antibiotic resistance not only through the digestive tract of animals but also in their final product such as milk. It is also possible the results show that human factor plays important role in the transfer resistant genes from human to animal.

REFERENCES

- BÍREŠ, J. 2004. Aktuálna legislatíva v oblasti hygieny mlieka. In *Mliekárstvo*, vol. 35, 2004, no. 1, p. 33-35.
- BOBKOVÁ, A., ZELENÁKOVÁ, L., LOPAŠOVSKÝ, L., PAVELKOVÁ, A. et al. 2008. Hodnotenie mikrobiologickej kvality smotanových jogurtov. In *Bezpečnosť a kontrola potravín: zborník prác z medzinárodnej vedeckej konferencie I. diel*. Nitra: Slovenská poľnohospodárska univerzita, 2008, p. 33-37.
- CARO, I., MATEO, J., GARCÍA-ARMESTO, M.R. 2007. Phenotypical characteristics of Shiga-like toxin *Escherichia coli* O157 isolated from sheep dairy products. In *Lett. Appl. Microbiol.* vol. 45, 2007 p. 295-300.
- ČÍZEK, A., DOLEJSKÁ, M., NOVOTNÁ, R., HAAS, D., VYSKOČIL, M. 2007. Survey of Shiga toxinogenic *Escherichia coli* O157 and drug-resistant coliform bacteria from in-line milk filters on dairy farms in the Czech Republic. In *J. Appl. Microbiol.* vol. 104, 2007, p. 852-860.
- DUPONT, H.L., WEST, H., EVANS, D.G., OLARTE, J., EVANS, D.J. 1978. Antimicrobial susceptibility of enterotoxigenic *Escherichia coli*. In *J. Antimicrob. Chem.*, vol. 4, 1978, p. 100-102.
- EFSA. 2008. Foodborne antimicrobial resistance as a biological hazard. In *Draft Scientific Opinion of the Panel on Biological Hazards* (Question No EFSA-Q-2007-089). Draft endorsed on 6 March 2008.
- EUCAST. 2010. European Committee on Antimicrobial Susceptibility Testing. Eucast disk diffusion test: breakpoint tables v1.1, April 27, 2010. Available on the internet: <http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Disk_test_documents/EUCAS_T_breakpoints_v1.1.pdf>.
- FARZANA, K., AKHTAR, S., JABEEN, F. 2009. Prevalence and antibiotic resistance of bacteria in two ethnic milk based products. In *Pak. J. Bot.*, vol. 41, 2009, no. 2, p. 935-943.
- KHARAZMI, Y., HAMMES, W.P., HERTELI, C. 2002. Construction of a marker rescue system in *Bacillus subtilis* for detection of horizontal gene transfer in food. In *Syst. Appl. Microbiol.*, vol. 25, 2002, no. 3, p.471-477.
- LADO, B., YOUSEF, A. 2002. Alternative foodpreservation technologies: efficacy a mechanisms. In *Microbes and Infection*. vol. 4, 2002, no. 6, p. 433-440.
- LIRA, W.M., MACEDO, C., MARIN, J.M. 2004. The incidence of Shiga toxin-producing *Escherichia coli* in cattle with mastitis in Brazil. In *J. Appl. Microbiol.* vol. 97, 2004, p. 861-866.
- Mc MAHON, M.A.S., BLAIR, I.S., MOORE, J.E., Mc DOWELL, D.A. 2007. The rate of horizontal transmission of antibiotic resistance plasmids is increased in food preservation-stressed bacteria. In *J. Appl. Microbiol.*, vol. 10, 2007, no. 3, p. 1883-1888.
- PICOZZI, C., FOSCHINO, R., HEUVELINK, A., BEUMER, R. 2005. Phenotypic and genotypic characterization of sorbitol-negative or slow-fermenting (suspected O157) *Escherichia coli* isolated from milk samples in Lombardy region. In *Lett. Appl. Microbiol.* vol. 40, 2005, p. 491-496.
- SOLOMAKOS, N., GOVARIS, A., ANGELIDIS, A.S., POURNARAS, S., BURRIEL, A.R., KRITAS, S.K., PAPAGEORGIOU, D.K. 2009. Occurrence, virulence genes and antibiotic resistance of *Escherichia coli* O157 isolated from raw bovine, caprine and ovine milk in Greece. In *Food Microbiology*, vol. 26, 2009, p. 865-871.

Acknowledgements: This study was supported by VEGA č.2/0012/08, VEGA č. 1/0372/09; KEGA č. 430-014SPU-4/2010.

Contact address: Ing. Lukáš Hleba, Slovak University of Agriculture in Nitra, Faculty of biotechnology and food science, Department of Microbiology, Tr.Andreja Hlinku 2, 949 76 Nitra, lukas.hleba@gmail.com